

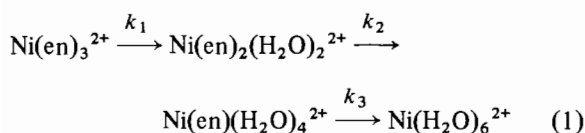
Visible Spectra of the Complex Species Ni(en)_3^{2+} , $\text{Ni(en)}_2(\text{H}_2\text{O})_2^{2+}$ and $\text{Ni(en)}(\text{H}_2\text{O})_4^{2+}$ from Kinetic Studies

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(Received October 3, 1986)

It has been shown by several authors [1–3] that acid hydrolysis of the complex cation Ni(en)_3^{2+} (en = 1,2-diamino-ethane) is a three-step process which follows reaction (1):



At 25 °C the following rate data were obtained [1]:

$$k_1 = 86.6 \text{ s}^{-1}; k_2 = 5.2 \text{ s}^{-1}; k_3 = 0.145 \text{ s}^{-1}$$

Applying a rapid-scanning spectrometer Moore *et al.* [3] reported values of λ_{max} (wavelength of maximum absorption) and ϵ_{max} (absorption coefficient at λ_{max}) for the species $\text{Ni(en)}_2(\text{H}_2\text{O})_2^{2+}$ and $\text{Ni(en)}(\text{H}_2\text{O})_4^{2+}$. These data were taken from the spectra obtained at intermediate stages of the hydrolysis process.

The present contribution presents data for λ_{max} and ϵ_{max} of the above mentioned species which were obtained by studying reaction (1) with a newly designed, high resolution rapid-scan-stopped-flow spectrophotometer [4] and by analyzing a total of 90 spectra with a computer program producing time-independent values of λ_{max} and ϵ_{max} for the intermediates.

Experimental

In the stopped-flow experiment equal amounts of a 0.10 M Ni(en)_3^{2+} solution (0.10 M in reagent grade $\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$ and 0.355 M in freshly distilled 'en', which corresponds to 18% excess of the ligand) and of 1.0 M HCl were mixed.

The set-up and the specifications of the rapid-scan-stopped-flow spectrophotometer have been described elsewhere [4]. In the stopped-flow experiment a total of 90 spectra were taken in the wavelength range 304–725 nm. The absorbance/time

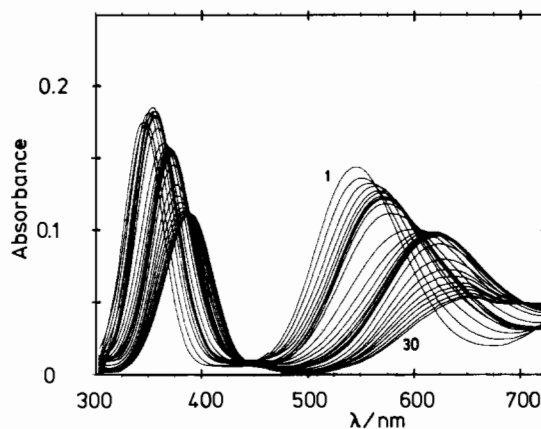


Fig. 1. Spectral changes observed for the various stages of reaction (1) at 5 °C with Δt (time interval between 2 consecutive spectra) being 30 ms (spectra 1–10), 600 ms (spectra 11–20) and 15 s (spectra 21–30).

data at 5 different wavelengths were fitted to function (2), which led to values for a_0 , a_1 , a_2 , a_3 and $A = a_0 + a_1 \exp(-k_1 t) + a_2 \exp(-k_2 t) + a_3 \exp(-k_3 t)$ (2)

averaged values for k_1 , k_2 , k_3 by an iteration procedure. On the basis of the averaged rate constants the amplitudes a_i for all wavelengths (*i.e.*, the complete spectra) were calculated. The spectra of all four species involved in reaction (1) were finally calculated on the basis of the amplitudes a_i .

Results and Discussion

Figure 1 presents every third spectrum out of a total of 90 spectra taken in a typical stopped-flow experiment. The spectra were taken at three different time bases, Δt (time interval between two consecutive spectra) being 10 ms in the beginning (spectra 1–29), 200 ms in the middle (spectra 30–59) and 5 s at the end (spectra 60–89). It is obvious that during the hydrolysis process the spectra are red-shifted, in both the wavelength range 350–400 nm (${}^3\text{T}_1(\text{P}) \leftarrow {}^3\text{A}_2$) and 550–650 nm (${}^3\text{T}_1(\text{F}) \leftarrow {}^3\text{A}_2$). This red-shift is accompanied by a loss in intensity for both absorption bands.

Computer fitting of the absorbance/time data to eqn. (2) at 5 different wavelengths leads to the rate constants listed in Table I. It follows that the rate of hydrolysis decreases with decreasing numbers of chelate ligands being coordinated, *i.e.*, $k_1:k_2:k_3 = 533:30:1$. The comparison with the rate constants reported in the literature [1, 3] suffers from the fact that the studies were carried out at different temperatures. Using the activation energies pre-

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TABLE I. Rate Constants for Reaction (1) at 5 °C

k_1 (s ⁻¹)	k_2 (s ⁻¹)	k_3 (s ⁻¹)	k_1/k_2	k_2/k_3	Reference
11.4 ± 1.0	0.645 ± 0.014	0.0214 ± 0.0011	17.7	30.1	this work
11	0.54	0.015	20.6	36.0	1 ^a
6.8	0.28	0.013	24.3	21.5	3 ^a

^aRate constants at 5 °C calculated from those at higher temperature with the activation energies given in ref. 1.

TABLE II. Wavelengths of Maximum Absorption (λ_{\max}) and Absorption Coefficients (ϵ_{\max}) of the Species Involved in Reaction (1)

Species	³ T ₁ (P) ← ³ A ₂		³ T ₁ (F) ← ³ A ₂		Reference
	λ_{\max} (nm)	ϵ_{\max} (nm)	λ_{\max} (nm)	ϵ_{\max} (M ⁻¹ cm ⁻¹)	
Ni(en) ₃ ²⁺	343.8 ± 1	7.7	542.8 ± 1	6.2	this work 3
	345 ± 5	7.8	550 ± 5	6.0	
Ni(en) ₂ (H ₂ O) ₂ ²⁺	356.4 ± 1	7.9	570.1 ± 1	5.2	this work 3
	360 ± 5	7.8	570 ± 5	5.0	
Ni(en)(H ₂ O) ₄ ²⁺	371.5 ± 1	6.6	620.5 ± 1	4.1	this work 3
	373 ± 5	6.7	625 ± 5	3.8	
Ni(H ₂ O) ₆ ²⁺	388.3 ± 1	5.3	651.8 ± 1	1.9	this work 3
	390 ± 5	5.1	650 ± 5	1.6	

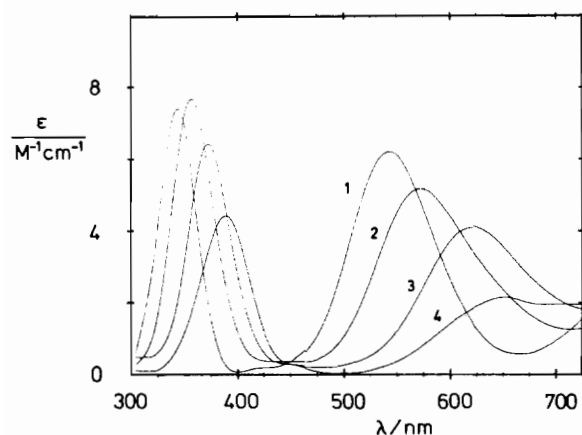


Fig. 2. Visible spectra of the species Ni(en)₃²⁺ (1), Ni(en)₂(H₂O)₂²⁺ (2), Ni(en)(H₂O)₄²⁺ (3) and Ni(H₂O)₆²⁺ (4) at 5 °C.

sented by Wilkins *et al.* [1] for converting all rate constants to 5 °C one obtains the data given in Table I. Considering the errors involved (the activation parameters have an error of more than 5%) the agreement is acceptable.

Computer approximation of the absorption coefficients ϵ of all four species involved in reaction (1) leads to the spectra shown in Fig. 2 and to the values of λ_{\max} and ϵ_{\max} listed in Table II. These values prove indeed the red-shift and the loss in intensity which accompanies the loss of coordinated chelate ligands, *i.e.*, the stepwise transition from the NiN₆ coordination to the NiO₆ coordination. Due to the fact that the resolution of the diode

array spectrophotometer used in this work (0.8 nm/diode) is much better than that of the set-up used by Moore *et al.* [3] the error in λ_{\max} is only ±1 nm. In addition, the way in which the spectra of the intermediates were calculated (see 'Experimental') excludes any contributions from other species. The agreement between the values for λ_{\max} and ϵ_{\max} obtained in this work and those obtained from intermediate spectra [3] is nevertheless sufficiently good.

The intermediate Ni(en)₂(H₂O)₂²⁺ could possibly exist in the *cis* or *trans* forms. It follows from the experimental evidence presented by Moore *et al.* [3] that the observed spectrum of the species Ni(en)₂(H₂O)₂²⁺ corresponds well with that of known *cis* octahedral species.

Acknowledgements

The authors thank the Deutsche Forschungsgemeinschaft and the Verband der Chemischen Industrie e.V. for support.

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